

**Isolation of 2-Deoxy-20-hydroxyecdysone and 3-Epi-2-deoxyecdysone
from Eggs of the Desert Locust, *Schistocerca gregaria*,
during Embryogenesis**

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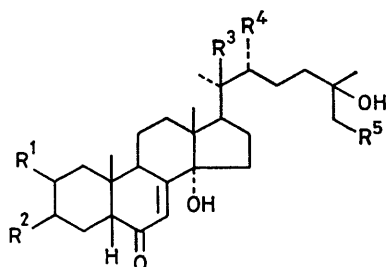
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Summary 2-Deoxy-20-hydroxyecdysone and 3-epi-2-deoxyecdysone have been isolated from developing eggs of the desert locust, *Schistocerca gregaria*, and their structures determined.

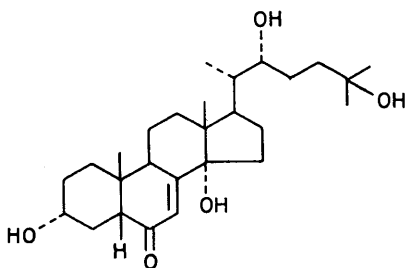
THE ovaries of several species of insects have been shown to synthesise ecdysteroids (insect moulting hormones), which

in most cases are passed largely into the eggs.¹ In the desert locust, *Schistocerca gregaria*, the ecdysteroids which have been identified^{2,3} in newly laid eggs include ecdysone (1), 2-deoxyecdysone (2), and 20-hydroxyecdysone (3), together with probable traces of 26-hydroxyecdysone (4), all being present primarily as polar conjugates hydrolysable with a crude *Helix pomatia* enzyme preparation. In this

insect, ecdysteroids are also present primarily as conjugates throughout embryogenesis,⁴ which generally lasts 17.5 days at 30 °C. The isolation and identification of 2-deoxy-20-hydroxyecdysone (5) and 3-epi-2-deoxyecdysone (6) from 17-day-old eggs of *S. gregaria* are now reported; (6) has not previously been isolated from insects.



- (1) $R^1=R^2=R^4=OH, R^3=R^5=H$
 (2) $R^2=R^4=OH, R^1=R^3=R^5=H$
 (3) $R^1=R^2=R^3=R^4=OH, R^5=H$
 (4) $R^1=R^2=R^4=R^5=OH, R^3=H$
 (5) $R^2=R^3=R^4=OH, R^1=R^5=H$



(6)

Day-17 developing eggs (160 g) of *S. gregaria* were extracted and fractionated as previously described.³ Ecdysteroids were released from the conjugate fraction by hydrolysis with *Helix pomatia* enzyme and fractionated directly by h.p.l.c. on a reversed-phase column (Partisil ODS-3).† Two u.v.-absorbing compounds [λ_{max} (in MeOH) 242 nm] were isolated [(5), 100 μ g; (6), 125 μ g] in addition to the ecdysteroids (1)–(4) previously identified in newly laid eggs.

The mass spectra of (5) included the following ions: chemical ionisation (c.i.) (isobutane) m/e , 465 ($[M + H]^+$, 6%); electron impact (e.i.) m/e , 428 ($[M - 2H_2O]^+$, 0.5%), 392 ($[M - 4H_2O]^+$, 2%), 347 ($[M - (C-22-C-27)]^+$, 9%), 284 ($[M - (C-20-C-27) - H_2O]^+$, 6%), 99 ($[C-22-C-27 - H_2O]^+$, 17%), 81 ($[99 - H_2O]^+$, 34%), and 43 (100%). The 1H n.m.r. spectrum (Fourier Transform, 400 MHz, C_5D_5N) of (5) showed signals at δ 1.08 (3H, s, 19-Me), 1.27 (3H, s, 18-Me), 1.40 (6H, s, 26/27-Me), and 1.64 (3H, s, 21-Me). Acetylation of (5) for 17 h at room temperature gave primarily a diacetate: c.i.-m.s. (NH_3), m/e , 513 ($[M +$

$H - 2H_2O]^+$, 0.1%), 495 ($[M + H - 3H_2O]^+$, 0.5%), 393 ($[513 - 2 \times 60]^+$, 2%), and 43 (100%).

The mass spectral data suggest that (5) is a pentahydroxylated ecdysteroid and are consistent with the presence of hydroxy-groups at C-20, C-22, and C-25. The methyl signals in the 1H n.m.r. spectrum of (5) were similar to those of 20-hydroxyecdysone and indicate the presence of hydroxy-groups at C-14, C-20, and C-25.⁵ From biogenetic considerations, the final hydroxy-group can probably be placed at C-3. These data taken together indicate that (5) is 2-deoxy-20-hydroxyecdysone. This was confirmed by comparison of (5) by t.l.c., h.p.l.c. (Partisil ODS-3 and APS-Hypersil)⁶ and e.i.-m.s. with an authentic sample supplied by Dr. D. H. S. Horn.

The mass spectra of (6) included the following ions: c.i. (isobutane) m/e , 449 ($[M + H]^+$, 1%; e.i., m/e , 430 ($[M - H_2O]^+$, 4%), 394 ($[M - 3H_2O]^+$, 4%), 343 ($[M - 5H_2O - Me]^+$, 5%), 332 ($[M - (C-22-C-27)]^+$, 8%), 284 ($[M - (C-20-C-27) - H_2O]^+$, 18%), 99 ($[C-22-C-27 - H_2O]^+$, 100%), and 81 ($[99 - H_2O]^+$, 68%); 1H n.m.r. (Fourier Transform, 400 MHz, C_5D_5N) δ 0.75 (3H, s, 18-Me), 1.01 (3H, s, 19-Me), 1.32 (3H, d, J 6 Hz, 21-Me), and 1.41 (6H, s, 26/27-Me).

The c.i. and e.i. spectra of (6) were almost indistinguishable from those of 2-deoxyecdysone. The methyl signals in the 1H n.m.r. spectrum of (6) were similar to those of ecdysone (1)⁵ except for the resonance of the C-19 methyl group. The chemical shift (δ 1.01, pyridine) of the C-19 methyl in (6) occurs at higher field than that of 2-deoxy-20-hydroxyecdysone (δ 1.08) and 2-deoxyecdysone (2).³ Such an up-field shift has been reported in 3-epi-2-deoxy-20-hydroxyecdysone as compared to 2-deoxy-20-hydroxyecdysone,⁷ but is not observed in 3-epi-ecdysteroids possessing a hydroxy-group at C-2.⁸

Acetylation of (6) for 17 h at room temperature gave primarily a diacetate: c.i.-m.s. (NH_3), m/e , 533 ($[M + H]^+$, 1%); 1H n.m.r. (Fourier Transform, 400 MHz, $CDCl_3$) δ 2.02 (3H, s, 3-OAc), 2.07 (3H, s, 22-OAc),⁵ 4.73 (1H, m, 3-H, $w_{1/2}$ 22 Hz), 5.87 (1H, d, 7-H), and 4.90 (1H, m, 22-H, $w_{1/2}$ 16 z). The broad resonance of the axial C-3 proton is in agreement with that reported for 3-epi-2-deoxy-20-hydroxyecdysone.⁷

These data taken together establish the structure of (6) as 3-epi-2-deoxyecdysone. This was corroborated by comparison of (6) by t.l.c., h.p.l.c. (Partisil ODS-3) and e.i.-m.s. with a sample of 3-epi-2-deoxyecdysone recently isolated from the fern, *Blechnum vulcanicum*.⁹

2-Deoxy-20-hydroxyecdysone is present throughout embryogenesis in *S. gregaria* eggs and is, at least primarily, of maternal origin. This ecdysteroid may be formed from 2-deoxyecdysone. The possibility exists that 20-hydroxyecdysone may arise not only by C-20 hydroxylation of ecdysone but also by hydroxylation of 2-deoxy-20-hydroxyecdysone at C-2. 2-Deoxy-20-hydroxyecdysone has been isolated previously from insects and crayfish.¹⁰ 3-Epi-2-deoxyecdysone is detectable only in developing *S. gregaria* eggs towards the end of embryogenesis (day 14, onwards). This ecdysteroid has not hitherto been reported in insects.

† Retention volumes on a Whatman Magnum 9 Partisil ODS-3 column (50 cm \times 9.4 mm i.d.) eluted with a linear gradient (40 min) 40 \rightarrow 80% methanol-water were: ecdysone (1), 110; 2-deoxy-20-hydroxyecdysone (5) 120; 3-epi-2-deoxyecdysone (6) 154 ml. R_f values on silica gel t.l.c. (solvent, $CHCl_3$ -MeOH 4:1) in comparison with other known ecdysteroids were: (6) 0.60; 2-deoxyecdysone (2) 0.56; (5) 0.51; ecdysone (1) 0.39.

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